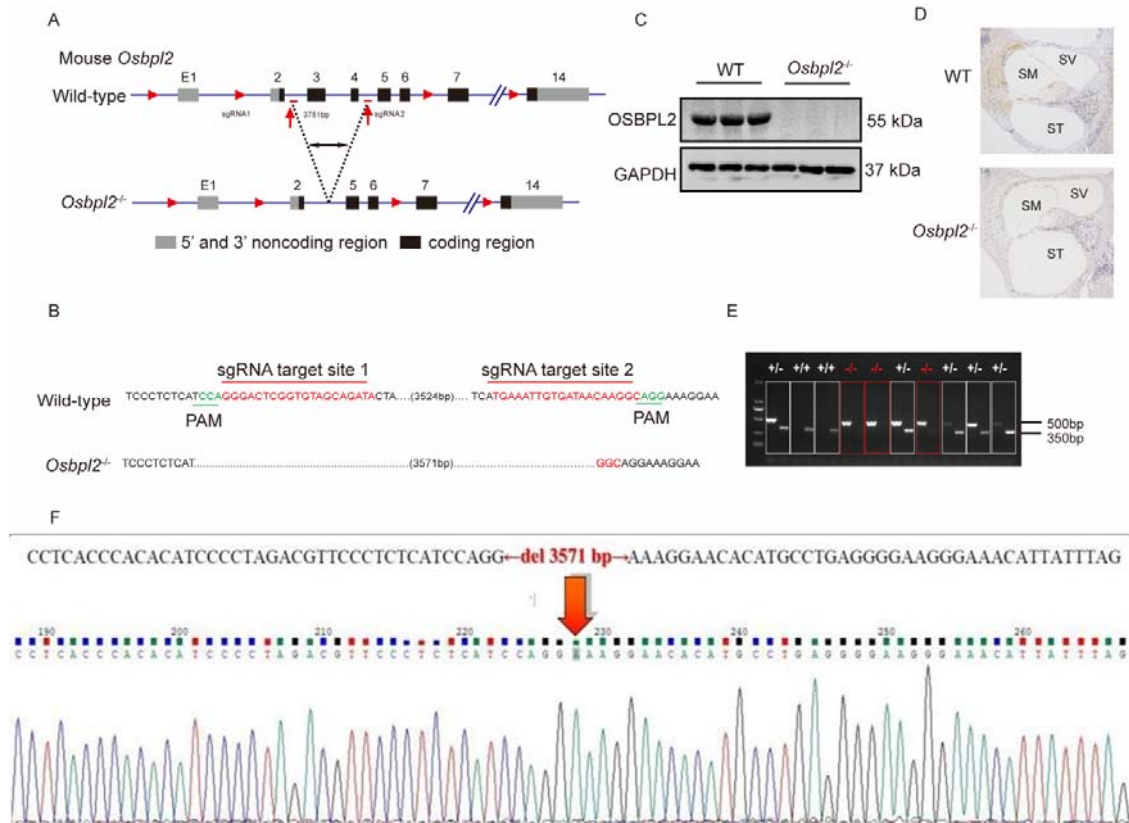
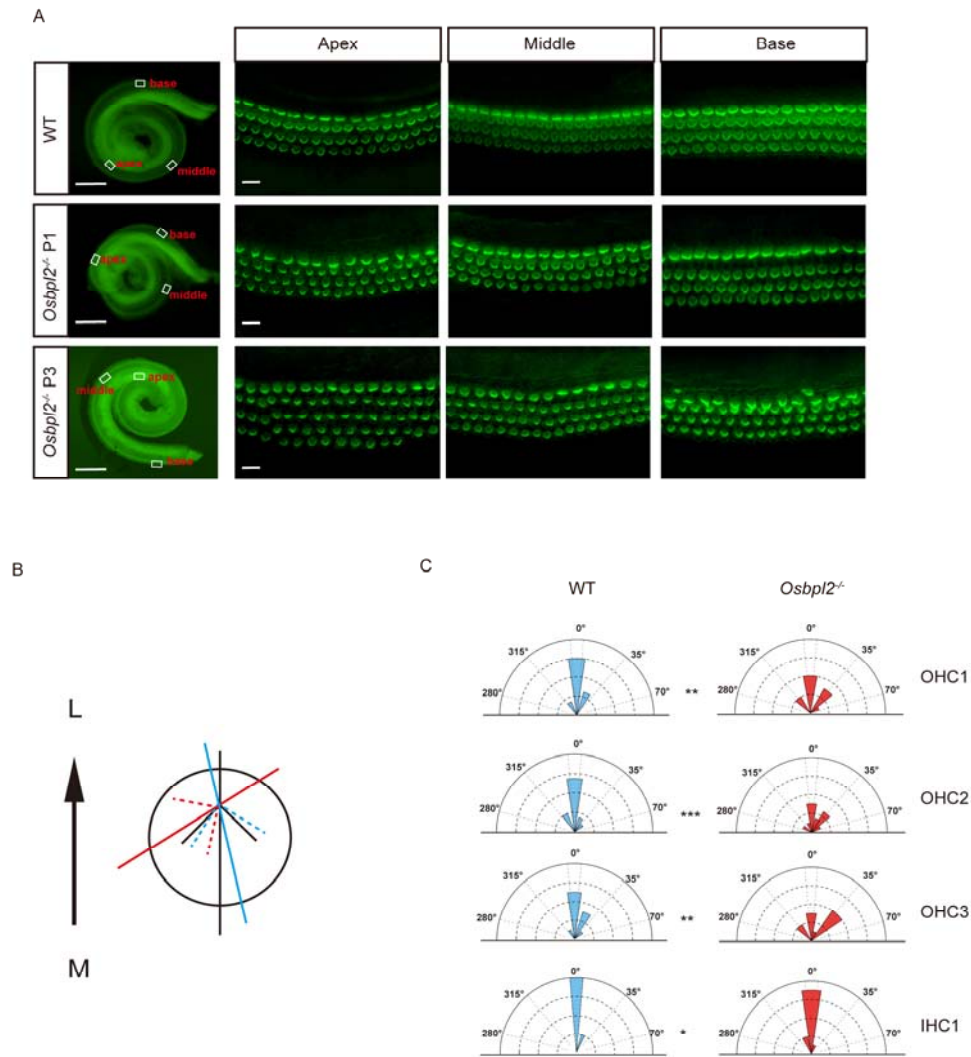


Supplemental Figures:

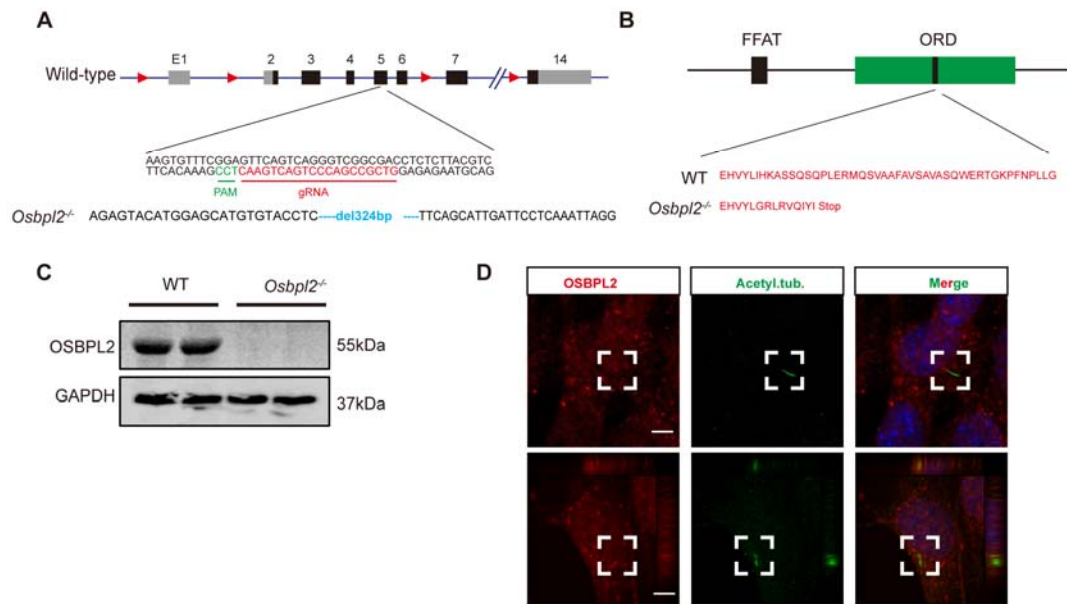


Supplemental Figure 1 Generation of *Osbpl2*^{-/-} mice. (A) Schematic diagram of mouse *Osbpl2* gene. (B) SgRNAs were designed to target the region flanking exons 3 and exon 4 in mouse *Osbpl2* gene. SgRNA sequence (red), PAM domain (green). (C) Western blot analysis of OSBPL2 in cochleae of 1-month-old *Osbpl2*^{-/-} mice (n=3). (D) Immunohistochemistry analysis of OSBPL2 in cochlea of 1-month-old *Osbpl2*^{-/-} and WT mice. SV: scala vestibuli; SM: scala media; ST: scala tympani. (E) PCR analysis of genomic DNA from the tail biopsy of *Osbpl2*^{-/-}, *Osbpl2*^{+/-} and WT mice. *Osbpl2*^{-/-}: 500 bp, *Osbpl2*^{+/-}: 500 bp / 350 bp, WT: 350 bp. (F) The *Osbpl2*^{-/-} mice carried a homozygous deletion of 3571 bp in mouse *Osbpl2* gene.

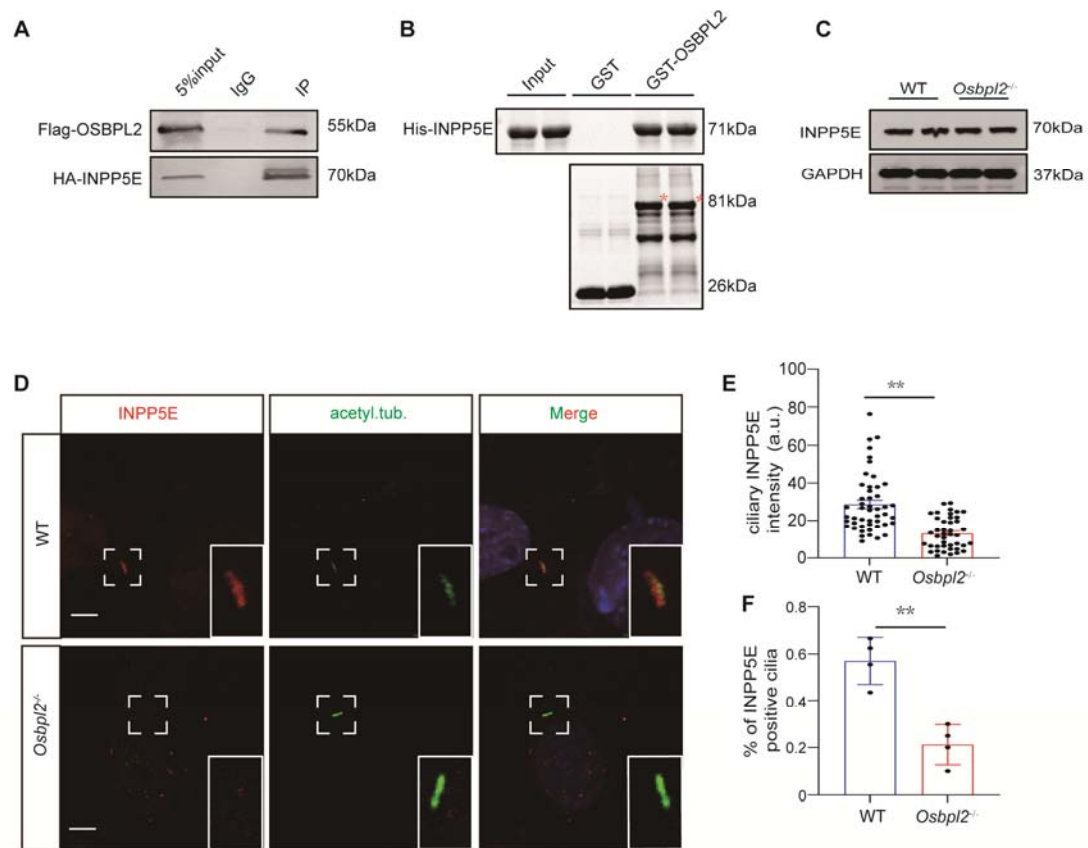


Supplemental Figure 2 Planar cell polarity (PCP) defects in *Osbp12*^{-/-} mice cochlea.

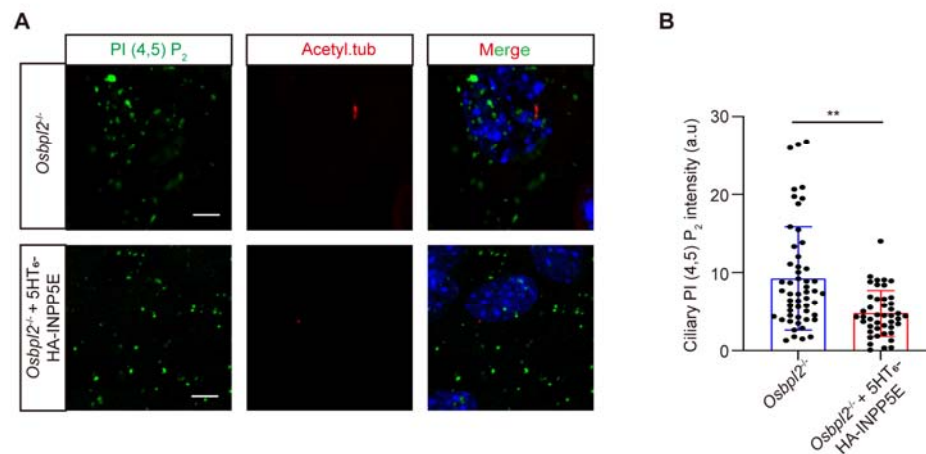
(A) Immunofluorescence staining of sensory epithelium in *Osbp12*^{-/-} and WT mice (P1 and P3) with phalloidin (green). Scale bar: 5μm. **(B)** Schematic diagram of stereocilia bundles polarity. The HCs polarity was evaluated by measuring the angle between the stereociliary bundles and the medio-lateral axis of the epithelium. **(C)** Distribution of HCs orientation. The angle of deviation between the stereociliary bundle and the medio-lateral axis was plotted in rose diagrams (pooled data were obtained from 10-15 individual cells per row of HCs for 3 mice per genotype, * $p < 0.05$, ** $p < 0.01$ and *** $p < 0.001$ tested by Wilcoxon rank-sum test).



Supplemental Figure 3 Generation of *Osbpl2*^{-/-} HEI-OC1 cells. (A) Schematic representation of the *Osbpl2*-knockout strategy. A biallelic mutation in *Osbpl2* gene (*Osbpl2*^{-/-}) was detected in a positive cell clone. (B) *Osbpl2*-knockout led to truncated protein. (C) Western blot analysis of OSBPL2 in *Osbpl2*^{-/-} HEI-OC1 cells. (D) Immunofluorescence staining of the ciliary OSBPL2 in HEI-OC1 cells (serum-starved for 24 h) with anti-OSBPL2 (red), anti-acetylated tubulin (green) and DAPI (blue).



Supplemental Figure 4 OSBPL2 regulated the ciliary location of INPP5E. (A) Co-IP assay verified the interaction of OSBPL2 with INPP5E. HEK293Ta cells transfected with Flag-OSBPL2 was used as negative control. (B) *E. coli* expressing His-INPP5E was used in GST pulldown assays, which verified the interaction of OSBPL2 with INPP5E. (C) Immunoblot analysis of INPP5E in whole cell lysate of *Osbpl2*^{-/-} and WT HEI-OC1 cells. GAPDH served as a loading control. (D) Immunofluorescence staining of the ciliary INPP5E in *Osbpl2*^{-/-} and WT HEI-OC1 cells (serum-starved for 24 h) with anti-INPP5E (red), anti-acetylated tubulin (green) and DAPI (blue). Dashed frames denote the locally zoomed regions in solid frames (bottom right). Scale bar: 5μm. (E) Quantification of ciliary INPP5E intensity in *Osbpl2*^{-/-} and WT HEI-OC1 cells (at least 30 cells from a microscope field, each dot represents a cell. ***p*<0.01 tested by Student's *t* test). (F) The proportion of INPP5E positive cilia in *Osbpl2*^{-/-} and WT HEI-OC1 cells (at least 30 cells from a microscope field, each dot represents the proportion of INPP5E positive cilia in a microscope field. ***p*<0.01 tested by Student's *t* test).



Supplemental Figure 5 The accumulation of PI(4,5)P₂ at the base of the cilia was alleviated by INPP5E expression. (A) Immunofluorescence staining of the ciliary PI(4,5)P₂ in *Osbp12*^{-/-} HEI-OC1 cells expressing 5HT₆-HA-INPP5E. Cells were stained with anti-acetylated tubulin (red), anti-PI(4,5)P₂ (green) and DAPI (blue). Scale bar: 5 μm. **(B)** Quantification of ciliary PI(4,5)P₂ intensity in *Osbp12*^{-/-} HEI-OC1 cells expressing 5HT₆-HA-INPP5E (at least 30 cells from a microscope field, each dot represents a cell. ***p* < 0.01 tested by Student's *t* test).

Supplemental Table Captions

Supplemental Table 1 Primers for PCR-based sequencing and qRT-PCR

Table 1 Primers for PCR-based sequencing and qRT-PCR

Sequence Name	Sequence (5' to 3')
HA- <i>Inpp5e</i> -F	GTGCCAGACTACGCAGGATCCATGCCATCCAAGTCAGCT
HA- <i>Inpp5e</i> -F	TTTAATAAGATCTGGTACCTCAGGACACGGTGCAAACCT
Flag- <i>ORP2</i> -F	CGACGATGATAAGTCCGGATCCATGAACGGGGAGGAAGA
Flag- <i>ORP2</i> -R	TTTAATAAGATCTGGTACC GTATATGTCTGGGCAGT
5HT ₆ -HA-F	ATGGTTCCAGAGCCCCGGC
5HT ₆ -HA-R	TCAGTTCATGGGGGAACC
Flag- Δ ORD-F	CGACGATGATAAGTCCGGATCCATGAACGGGGAGGAAGAAT
Flag- Δ ORD-F	TTTAATAAGATCTGGTACCTTAACTTTTGCTGGTATCCAAGT
Flag- Δ FFAT-F	CGACGATGATAAGTCCGGATCC ATGACTAGGA
Flag- Δ FFAT-F	TTTAATAAGATCTGGTACC GTATATGTCTGGGCAGT
q-M- <i>Gli1</i> -F	CCAAGCCAACTTTATGTCAGGG
q-M- <i>Gli1</i> -R	AGCCCGCTTCTTTGTTAATTTGA
q-M- <i>Ptch1</i> -F	AAAGAACTGCGGCAAGTTTTTG
q-M- <i>Ptch1</i> -R	CTTCTCCTATCTTCTGACGGGT
q-M- <i>Gapdh</i> -F	AGGTCGGTGTGAACGGATTTG
q-M- <i>Gapdh</i> -R	TGTAGACCATGTAGTTGAGGTCA
<i>Osbpl2</i> -OC1-F	CAAATCGGAACCTGGAGGAAC
<i>Osbpl2</i> -OC1-R	CCCCGGCCTGGTACACTG
<i>Osbpl2</i> -mice-F1/F2	TATAGTGGACTTACCGAAATCTCAG
<i>Osbpl2</i> -mice-R1-KO	TGGCTACTAGACACTCCATTTC
<i>Osbpl2</i> -mice-R2-WT	CATCAATAAGGCTGAAGCCATC