

Supplemental Data

Reduced CCL/Be-specific CD4⁺ T cells in CCL3-deficient or peptide-MHC II CAR-T cell-treated mice

Michael T. Falta^{1#}, Masoom Raza^{1#}, Caley J. Nevienski¹, Tonya Brunetti², Rui Fu³, Rebecca M. Tucker¹, Joseph M. Gaballa¹, Faiz Minhajuddin¹, Kibrom M. Alula¹, Alberto Dinarello¹, Douglas G. Mack¹, Allison K. Martin¹, Joseph Onyiah¹, Michael Yarnell^{4,5}, Prashanth Francis⁴, Terry J. Fry^{4,5}, Lisa A. Maier^{1,6}, Andrew P. Fontenot¹, Charles A. Dinarello¹, Shaikh M. Atif^{1*}

¹ Department of Medicine, University of Colorado Anschutz Medical Campus, Aurora, CO 80045, United States of America

² Department of Immunology and Microbiology, University of Colorado Anschutz Medical Campus, Aurora, CO 80045, United States of America

³ New York Genome Center, New York, NY 10013, United States of America

⁴ Department of Pediatrics, University of Colorado Anschutz Medical Campus, Aurora, CO 80045, United States of America

⁵ Center for Cancer and Blood Disorders, Children's Hospital Colorado, Aurora, CO 80045, United States of America

⁶ Department of Medicine, National Jewish Health, Denver, CO 80206, United States of America

MTF and MR contributed equally to this work.

Supplementary Figure Legends

Supplementary Figure 1. Chemokine gene expression in the BAL fluid of BeO/LPS-treated HLA-DP2 Tg WT and CCL3^{-/-} mice. Mice were intratracheally treated with 3 doses of BeO (100 µg) on days 0, 1, and 2 and a single dose of LPS (10 µg) on day 2. BAL was collected from individual mice on day 3. **(A)** Quantitative PCR (q-RT-PCR) shows fold gene expression of the *Ccl3* and *Ccl4* genes relative to *Gapdh* in the lungs of PBS- or BeO/LPS-treated mice. **(B)** CCL3 and CCL4 protein levels were measured in the BAL fluid by ELISA. The data are representative of the (mean ± SEM) of two independent experiments. 1-way ANOVA was used to determine statistical significance. *P < 0.01, ***P < 0.001, ****P < 0.0001.

Supplementary Figure 2. Control ELISPOT assay. **(A)** HLA-DP2 Tg WT and CCL3^{-/-} mice were exposed to PBS or BeO as described in the Materials and Methods. Lung CD4⁺ T cells were purified on day 21 and the treatment and control groups were stimulated *ex vivo* with phytohemagglutinin (1 µg/ml). The ELISPOT data plot shows T cell secretion of IFN-γ represented as spot-forming units (SFUs) per 100,000 purified CD4⁺ T cells stimulated overnight. Data represent the mean ± SEM across combined experiments.

Supplementary Figure 3. IFN-γ and IL-17A-secreting cells are reduced in BeO-exposed CCL3^{-/-} mice on day 12. HLA-DP2 Tg WT and CCL3^{-/-} mice were exposed to BeO (100 µg) on days 0, 1, and 2 and treated with LPS (10 µg) on day 8. On day 12, mice were sacrificed 2 minutes after intravascular administration of anti-CD45 mAb. **(A)** Schematic design of the experiment. **(B)** Graph plots show the total number of cells in the lungs of PBS- or BeO/LPS-treated mice. **(C)** Frequency (left) and number (right) of CD4⁺ T cells in the lungs were determined on day 21. **(D)** Number of activated (CD44⁺) CD4⁺ T cells. **(E-F)** ELISPOT responses for IFN-γ **(E)** and IL-17A **(F)** as spot-forming units (SFUs) per 25,000 purified CD4⁺ T cells stimulated with BeSO₄ (100 µM). **(G)** Representative CCR7 expression on gated lung tissue-resident CD4⁺ T cells. Solid histograms show unstained cells in grey, PBS-treated WT in yellow, and CCL3^{-/-} in peach, BeO-treated WT in brown, and CCL3^{-/-} in purple. **(H)** Cumulative frequency of CCR7⁺ CD4⁺ T cells in the lungs of BeO-exposed mice. Error bars represent the mean ± SEM. 1-way ANOVA determined statistical significance. *P < 0.05, **P < 0.01, ***P < 0.001, ****P < 0.0001.

Supplementary Figure 4. Enhanced CD4⁺ T cell responses in *A. nidulans* exposed HLA-DP2 Tg WT and CCL3^{-/-} mice. Both WT and CCL3^{-/-} mice were exposed to *A. nidulans* (1x10⁸) spores/conidia on day 0, and the mice were sacrificed on day 14. A fluorescently labeled anti-CD45 mAb was injected intravenously 2 minutes before sacrifice to distinguish tissue-specific cells from circulating cells. On day 14, **(A)** flow cytometry plots show the frequency of CD4⁺ T cells in the lungs of PBS or *A. nidulans*-exposed mice. **(B)** Graph plots show the frequency and number of CD4⁺ T cells in the lungs of *A. nidulans*-exposed mice. Data are representative of the mean ± SEM of two independent experiments. 1-way ANOVA was used to determine statistical significance. *P < 0.05, **P < 0.01.

Supplementary Figure 5. Tetramer specificity validation. HLA-DP2-CCL4/Be and HLA-DP2-CLIP tetramer staining of (A) T cell hybridoma 8845-c3 that expresses a human TCR specific for the CCL4/Be neoantigen and restricted by HLA-DP2, and (B) gated CD4⁺ T cells from the lungs of HLA-DP2 Tg mice, exposed to BeO under the standard regimen. Both tetramers were used at a final concentration of 20 µg/ml.

Supplementary Figure 6. Phenotypic and functional analysis of BeO-exposed lung tissue-resident CD4⁺ T cells. HLA-DP2 Tg FVB/N (WT) mice were intratracheally exposed to BeO (100 µg) on days 0, 1, 2, 14, 15, 18, and 19 and a single dose of LPS (10 µg) on day 14. On day 21, mice were injected intravenously with anti-CD45 mAb (5 µg/mouse) 2 minutes before sacrifice. Single cells were prepared from the lungs and stimulated with BeSO₄ (75 µM) for 4h in the presence of Brefeldin A (1X). The cells were stained with HLA-DP2-CCL4/Be tetramer (20 µg/ml) for 2h as described in the Materials and Methods. Subsequently, the cells were surface-stained, followed by intracellular staining to detect functional molecules. (A) Flow cytometric plots show cell surface expression of PD-1, CD44, and CD62L molecules and intracellular staining of proliferation marker, Ki-67, and the functional molecules, IFN-γ, TNF-α, and IL-17A. Histogram colors represent FMO control (grey), CD44⁻Tet⁻ CD4⁺ T cells (DN, green), CD44⁺Tet⁻ CD4⁺ T cells (Tet⁻, Red), and CD44⁺Tet⁺ CD4⁺ T cells (Tet⁺, blue). (B) Bar graphs show the mean fluorescence intensity (MFI) of cell-surface and intracellular molecules for the various CD4⁺ T cell subsets in the lungs of BeO-exposed mice. Data are representative of two independent experiments. 1-way ANOVA was used to determine statistical significance. *P < 0.05, **P < 0.01, ***P < 0.001, ****P < 0.0001.

Supplementary Figure 7. BeO-exposure induced single or dual specificity antigen-specific (CCL3 and 4) CD4⁺ T cells in the lungs. HLA-DP2 Tg mice (n=10) were sensitized and boosted with BeO (100 µg) on days 0, 1, 2, 14, 15, 18, and 19 and a single dose of LPS (10 µg) on day 14. On day 21, lung cells were stained with HLA-DP2-CCL3/Be and HLA-DP2-CCL4/Be tetramers labeled with different fluorophores. (A) Pseudocolor flow cytometry plots show representative DP2-CCL3/Be (left), and DP2-CCL4/Be (right) tetramer binding of CD4⁺CD44⁺ lung T cells. (B) Representative HLA-DP2 tetramer co-staining of lung CD4⁺ T cells from BeO/LPS-exposed mice. (C) Summary of the overall percentage of lung CD4⁺ T cells that bind individual tetramers, or are cross-reactive, i.e., bind both CCL/Be tetramers.

Supplementary Figure 8. Presence of CD4⁺ T cells in the lungs of BeO/LPS-exposed mice treated with or without CAR-T cells. HLA-DP2 Tg mice were exposed to BeO on days 0, 1, 2, 14, 15, 18, and 19, and LPS (10 mg/ml) on day 14. Treated mice were given PBS or adoptively transferred with either control, DR-α-CAR-T cells, or CCL4/Be-CAR-T cells on day 8. Mice were sacrificed and examined on day 21. The frequency (left) and number (right) of CD4⁺ T cells in the lungs of exposed mice are shown. Data are shown as mean ± SEM, pooled from two independent experiments. 1-way ANOVA was used to determine statistical significance. ***P < 0.001, ****P < 0.0001.